

Heavy Metal Concentration in Plant Leaves from Three Sites in Mie Prefecture

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Abstract

To identify heavy metal accumulator plants suitable for phytoremediation, concentration of heavy metals was analyzed in leaves of woody and herbaceous plants obtained from three sites in Mie Prefecture. The elemental concentrations varied considerably among the different plant species. A large intraspecific variation was also observed in the elemental concentrations at the different sites, corresponding with the concentration of each element in the soil of the sites. *Chengiopanax sciadophylloides* was identified as a strong and specific Mn accumulator. In contrast, *Clethra barbinervis* seems to absorb many kinds of heavy metals from soil, but its ability to transfer a heavy metal from soil to plant was found to be lower than that of *C. sciadophylloides*.

Key Words: heavy metal concentration, hyperaccumulator, *Chengiopanax sciadophylloides*,
Clethra barbinervis

Introduction

Application of heavy metal-accumulating plants is a promising method for remediation of soils polluted with heavy metals. About 450 hyperaccumulator plant species for seven heavy metals (Cd, Co, Cu, Mn, Ni, Tl, and Zn) are known¹⁾.

Little research has been conducted in Japan on heavy metal-accumulating plants. The concentrations of various elements in wild plants grown at an area artificially polluted by industrialization were determined by Yamada et al.²⁾. Tatekawa et al.³⁾ reported high levels of Cd in few species of Asteraceae, Polygonaceae, Cyperaceae, and Liliaceae families. Memon et al.⁴⁻⁶⁾ analyzed heavy metal concentration in plants distributed in the temperate forest of central Japan. Furthermore, Harada and Hatanaka⁷⁾ analyzed natural background levels of trace elements in wild plants, whereas Takada et al.^{8,9)} examined heavy metal concentrations in a number of plants collected from many places in Japan, including polluted and non-polluted areas. Regarding Japanese heavy metal hyperaccumulators, Mizuno et al.¹⁰⁾ reported Ni accumulation by *Thlaspi japonicum* and Kubota and Takenaka¹¹⁾ identified *Arabis gemmifera* as a Cd hyperaccumulator.

In the present study, in order to locate and identify suitable plants for heavy metal accumulation, we analyzed concentration of heavy metals in leaves of woody and herbaceous plants obtained from three sites in Mie prefecture. In general, heavy metal hyperaccumulator plants grow in zones with high concentrations of heavy metals. Since serpentine soils and mineral belt areas are known to have high concentrations of some heavy metals, we also analyzed plants growing in areas with such soils in Mie prefecture.

Materials and Methods

Leaves of numerous woody and herbaceous plants that are common at the sampling site were collected from Site A (a forest at Shiraki-cho, Toba city, a serpentine soil area), Site B (Mt. Fujiwara, at Fujiwara-cho, Inabe city, a mineral belt zone), and Site C (Hirakura Experiment forest, Mie University at Misugi-cho, Tsu city). Mature intact leaves were collected from healthy branches or twigs of the plant. Soil samples (5-10 cm in depth) near each sampled plant in each site were collected.

The plant samples were washed with distilled and deionized water, dried at 70°C in an oven for 72 h, and then ground using mortar and pestle. The soil samples were air-dried for 1 to 2 weeks and then dried at 100°C for 2 days. The dried soil samples were passed through a 2 mm mesh sieve. Powdered plant leaves (0.1-1 g) were placed in plastic tubes with a plastic stopper. Twenty times (vol/wt) of 1 M HCl was added to the tubes, shaken reciprocally for 5 h, and filtered¹²⁾. One gram of each soil sample was placed in a tall beaker and digested with HNO₃ and HClO₄, then filtered and diluted with water to make the final volume 50 mL. The concentration of metals in the extracts and digested solutions was measured using an atomic absorption spectrometer (Shimadzu AA-6500).

Results and Discussion

Heavy metal concentrations in the soil samples from the three sites are shown in Table 1. Considerable variation was observed in the metal profiles in the soils from the three sites. The soil at Site A is a serpentine soil; high concentrations of Cr, Mn, Co, and Ni were detected in this soil. The soil at Site B is mineral belt soil; high concentrations of Cu, Zn, Cd, and Pb were detected in this soil. The soil at Site C has no special properties of heavy metal concentration.

Heavy metal concentrations in the plant samples are shown in Table 2. Because the elemental concentrations varied considerably among the plant species, we speculated that the mineral elemental status of each species might be a hereditary character under genetic control. Some intraspecific variations in elemental concentrations were observed at each site and a large intraspecific variation was observed among the different sites, depending on the concentration of elements in the soil of each site.

As shown in Tables 1 and 2, Mn concentration in *C. sciadophylloides* (range, 4,870-1,600 mg kg⁻¹; average, 3,106 mg kg⁻¹) at Site C was about 9 times that in soil (average 341 mg kg⁻¹); however, Mn concentration in other plants collected from site C did not demonstrate higher concentrations of heavy metals than that in soil except for *C. barbinervis* (range, 1,230-98 mg kg⁻¹; average 564 mg kg⁻¹) and *Wisteria floribunda* (408 mg kg⁻¹). Manganese concentration in *C. sciadophylloides* was the highest of all plants collected from Site C. Manganese concentration in *C. sciadophylloides* grown in a forest at Uresino-cho, Matsusaka city was over 10 g kg⁻¹¹³⁾. Because of its ability to selectively accumulate Mn, and the concentration of other heavy metals is not high, *C. sciadophylloides* is useful for phytomining of Mn.

Table 1 Heavy metal concentration of soils at three sites in Mie prefecture

Sites	Cr	Mn	Fe	Co	Ni	Cu	Zn	Cd	Pb
Site A	1990±750	1300±420	82600±13500	118±23	1310±200	10.6±5.7	78.4±21.2	0.00±0.00	6.6±3.7
Site B	39.4±10.4	513±261	47100±6000	6.54±3.40	24.5±7.21	70.6±21.9	222±45	0.62±0.56	121±38
Site C	10.8±3.1	341±235	34800±3400	5.22±2.61	7.3±0.9	10.7±1.0	102±76	0.00±0.00	23.4±7.0
Site A, Toba city		Site B, Mt. Fujiwara		Site C, University forest					

Table 2 Heavy metal concentrations in the leaves of each plant species

plant species	sampling site	mg kg ⁻¹										
		Cr	Mn	Fe	Co	Ni	Cu	Zn	Cd	Pb	Mg	Ca
<i>Abelia spatholata</i>	Site A	tr	31	115	tr	17.6	4.83	10.6	tr		9520	6230
<i>Abies firma</i>	Site C	tr	141	45	tr	1.31	1.76	15	tr	tr	1700	3030
<i>Acer carpinifolium</i>	Site C	tr	197	121	tr	1.3	13.6	73.9	tr	tr	5670	13400
<i>Acer palmatum</i>	Site B	tr	411	87	tr	2.43	6.18	44.3	0.54	1.61		
<i>Acer sieboldianum</i>	Site C	1.56	94		tr	0.59	3.51	36.1	tr	2.2		
<i>Acorus gramineus</i>	Site A	tr	67		tr	8.43						
<i>Aesculus turbinata</i>	Site C	tr	78		tr	0.83	5.59	11.8	tr	3.47		
<i>Albizia Julibrissin</i>	Site A	tr	29	59.9	tr	14.2	1.95	7.83	tr		7800	2390
<i>Alnus firma</i>	Site A	tr	59	99.4	tr	8.47	3.63	14.6	tr		5260	10100
<i>Alnus japonica</i>	Site C	2.23	156		tr	1.29	7.16	27.9	0.13	3.64		
<i>Angelica Miqueliana</i>	Site A	tr	123		tr	10.3						
<i>Aquilegia Buergerina</i>	Site A	tr	59		tr	33.9						
<i>Aralia elata</i>	Site A	tr	225	86.4	tr	28.6	6.55	28.9	tr		14700	5040
<i>Artemisia vulgaris</i>	Site A	tr	256		tr	13.9						
<i>Aucuba japonica</i>	Site B		216				5.87	30.6	0.58	6.32		
<i>Aucuba japonica</i>	Site B		346				4.44	22.1	0.51	3.03		
<i>Aucuba japonica</i>	Site B		1157				4.04	21.9	tr	tr		
<i>Benzoin umbellatum</i>	Site C	tr	335	52.3	tr	2.69	7.81	49.3	tr	tr	3580	4500
<i>Benzoin umbellatum</i>	Site A	tr	164	172	1.44	12	3.85	33.7	tr		8940	5670
<i>Buxus microphylla</i>	Site B		120				3.21	74.2	tr	tr		
<i>Callicarpa mollis</i>	Site A	tr	423	89.9	tr	4.97	7.09	23.4	tr		2860	5510
<i>Camellia japonica</i>	Site C	tr	226	40.9	tr	0.44	5.22	8.39	tr	tr	1150	8910
<i>Camellia japonica</i>	Site A	tr	1460		tr	10.1						
<i>Camellia japonica</i>	Site A	tr	1050		tr	10.7						
<i>Cardamine flexuosa</i>	Site A	tr	94		tr	39.9						
<i>Carpinus japonica</i>	Site C	tr	163	39.8	tr	0.85	5.9	13	tr	tr	2780	5620
<i>Carpinus japonica</i>	Site C	0.26	115		tr	0.75	0.77	11.3	tr	tr		
<i>Carpinus laxiflora</i>	Site C	1.72	198		tr	1.12	8.21	20	tr	2.64		
<i>Carpinus Tschonoskii</i>	Site C		93		tr	0.88	9.69	55.7	tr	1.78		
<i>Celtis sinensis</i>	Site A	tr	59	91.9	tr	7.11	7.47	11	tr		13400	24300
<i>Cephalotaxus drupacea</i>	Site A		231		tr	19.72						
<i>Cephalotaxus drupacea</i>	Site A		170		tr	25.7						
<i>Chengiopanax sciadophylloides</i>	Site C	1.08	4870		tr	1.53	5.68	15	0.05	2.79		
<i>Chengiopanax sciadophylloides</i>	Site C	1.69	4380		tr	2.66	6.4	27.3	0.09	2.61		
<i>Chengiopanax sciadophylloides</i>	Site C	3.51	2390		tr	2.22	4.84	34.5	0.13	3.45		
<i>Chengiopanax sciadophylloides</i>	Site C	2.9	2290		tr	3.16	7.53	80.3	0.18	3.1		
<i>Chengiopanax sciadophylloides</i>	Site C	tr	1600	63.1	tr	3.72	14.4	27.6	tr	tr	8880	4590
<i>Cinnamomum Camphora</i>	Site A	tr	223	42.4	tr	3.57	9.71	13.2	tr		4930	3050
<i>Cinnamomum japonicum</i>	Site C	tr	43	92.6	tr	tr	3.22	7.03	tr	tr	780	5540
<i>Cinnamomum japonicum</i>	Site A	tr	212		tr	60.1						
<i>Cinnamomum japonicum</i>	Site A	tr	109	73.3	tr	24.5	3	8.75	tr		3620	4570
<i>Cinnamomum japonicum</i>	Site A	tr	58		tr	39.4						
<i>Cinnamomum japonicum</i>	Site A	tr	101		tr	54.7						
<i>Cinnamomum japonicum</i>	Site A	tr	118		tr	88.2						
<i>Cinnamomum japonicum</i>	Site B		57				5.32	9.22	tr	tr		
<i>Cinnamomum japonicum</i>	Site A	tr	221		tr	84						
<i>Cirsium japonicum</i>	Site A		156		tr	88.03						

plant species	sampling site	Cr	Mn	Fe	Co	Ni	Cu	Zn	Cd	Pb	Mg	Ca
<i>Cirsium japonicum</i>	Site A		89		tr	9.3						
<i>Cirsium japonicum</i>	Site A		59		tr	62.7						
<i>Citrus Unshiu</i>	Site B	1.7	11		0.2	0.42	5.37	13.5	0.01	2.3		
<i>Clethra barbinervis</i>	Site C	1.72	689		12.3	11	5.05	238	0.99	3.28		
<i>Clethra barbinervis</i>	Site C	2.05	1230		42.8	12.9	5.48	421	2.36	4.11		
<i>Clethra barbinervis</i>	Site C	1.56	736		17.7	8.33	5.38	301	1.16	3.83		
<i>Clethra barbinervis</i>	Site C	1.81	500		12.9	8.81	5.28	295	1.1	3.05		
<i>Clethra barbinervis</i>	Site C	2.32	338		1.51	3.89	6.98	11.6	0.67	3.2		
<i>Clethra barbinervis</i>	Site C	tr	98	50.2	4.76	8.02	13.8	132	tr	tr	4150	1210
<i>Clethra barbinervis</i>	Site C	2.2	619		2.6	4.95	5.52	79.6	0.47	3.16		
<i>Clethra barbinervis</i>	Site C	1.22	304		2.23	3.1	5.51	61	0.46	3.13		
<i>Clethra barbinervis</i>	Site A	tr	1220	190	295	110	6.22	116	0.99		14600	1790
<i>Clethra barbinervis</i>	Site A	2.9	1200		25.4	223	6.35	96.9	0.56	3.21		
<i>Clethra barbinervis</i>	Site A	8.28	1140		25.1	230	3.97	186	0.92	3.26		
<i>Clethra barbinervis</i>	Site A	tr	1100	60.5	21	104	4.01	15.6	tr		13800	6790
<i>Clethra barbinervis</i>	Site A	1.98	677		122	97.5	6.78	93.4	0.8	2.71		
<i>Clethra barbinervis</i>	Site A	1.5	1070		279	211	4.32	87	0.68	2.71		
<i>Clethra barbinervis</i>	Site A	tr	560	203	39.4	150	4.99	67.8	0.47		14000	6490
<i>Clethra barbinervis</i>	Site A	2.48	1490		140	332	6.27	90.9	0.75	3.06		
<i>Clethra barbinervis</i>	Site A	1.68	1050		158	278	6.6	89.5	0.58	2.56		
<i>Clethra barbinervis</i>	Site A	2.26	470		43.3	407	4.08	66.9	0.72	3.1		
<i>Clethra barbinervis</i>	Site A	2.36	1210		174	290	4.73	136	1.24	2.58		
<i>Clethra barbinervis</i>	Site A	2.11	496		19.3	158	7.59	95.6	0.5	3.23		
<i>Clethra barbinervis</i>	Site A	1.26	725		60.9	167	4.6	110	0.45	2.72		
<i>Clethra barbinervis</i>	Site B	tr	1530	110	58.7	33.1	5.43	246	10.9	1.43		
<i>Clethra barbinervis</i>	Site B	0.14	1590		69.3	43.3	6.87	288	10	1.62		
<i>Clethra barbinervis</i>	Site B	tr	1050		23.8	17.3	7.49	323	18.6	2.3		
<i>Clethra barbinervis</i>	Site B	tr	1030	103	15.2	13	7.73	275	14.6	16.3		
<i>Clethra barbinervis</i>	Site B	0.35	604		28.5	13.8	9.14	354	8.23	1.65		
<i>Clethra barbinervis</i>	Site B	tr	1130	80.4	31.8	25.3	6.58	227	6.98	2.64		
<i>Clethra barbinervis</i>	Site B	0.48	1220		34.3	25	10.8	278	7.28	1.8		
<i>Clethra barbinervis</i>	Site B	1.13	1360		59.8	23	8.29	196	6.29	1.8		
<i>Clethra barbinervis</i>	Site B	tr	1090	81.2	24.5	31	5.3	179	5.42	1.43		
<i>Clethra barbinervis</i>	Site B	0.3	1230		108	13	8.32	74.4	2.09	1.65		
<i>Clethra barbinervis</i>	Site B	0.61	2010		45.2	50.1	8.47	383	5.48	1.02		
<i>Clethra barbinervis</i>	Site B	0.42	459		12.6	15.3	5.79	187	3.35	1.05		
<i>Clethra barbinervis</i>	Site B	tr	2230	110	35.5	21.9	5.81	294	6.84	tr		
<i>Clethra barbinervis</i>	Site B	0.1	1690		37	42.5	5.15	196	6.46	1.7		
<i>Clethra barbinervis</i>	Site B	tr	928	82.8	56.7	13.2	8.79	97.4	22.3	tr		
<i>Clethra barbinervis</i>	Site B	tr	1170	79.7	14.6	16.8	5.82	269	4.57	tr		
<i>Cleyera ochracea</i>	Site C	tr	54	35.9	tr	7.81	3.77	8.71	tr	tr	2280	7870
<i>Cleyera ochracea</i>	Site A	tr	206	154	tr	14.2	1.52	6.1	tr		13900	13900
<i>Cleyera ochracea</i>	Site A	tr	345		tr	17.3						
<i>Cleyera ochracea</i>	Site A	tr	243		tr	16						
<i>Cornus controversa</i>	Site C	tr	111	50.6	tr	0.45	6.57	74.3	0.31	tr	2510	4690
<i>Cryptomeria japonica</i>	Site A	tr			tr	7.47						
<i>Cryptomeria japonica</i>	Site A	tr			tr	3.81						
<i>Damanacanthus indicus</i>	Site A		42		tr	49.19						
<i>Daphniphyllum macropodum</i>	Site B		19				3.91	17.4	tr	tr		

plant species	sampling site	Cr	Mn	Fe	Co	Ni	Cu	Zn	Cd	Pb	Mg	Ca
<i>Daphniphyllum macropodum</i>	Site B		17				2.72	24.7	tr	3.35		
<i>Deutzia Maximowicziana</i>	Site C	tr	54	58.7	tr	2.09	3.29	14.6	tr	tr	8040	36600
<i>Diospyros Kaki</i>	Site A	tr	295	85.8	tr	9.47	1.58	69.2	0.82		11400	2660
<i>Elaeagnus glabra</i>	Site A	tr	49		tr	10.2						
<i>Elaeagnus pungens</i>	Site B	tr	187	57.8	tr	7.39	14.7	19.6	tr	tr		
<i>Elaeagnus pungens</i>	Site B		1340				11.7	29.6	tr	3.41		
<i>Elaeagnus pungens</i>	Site B		251				13.7	26.8	tr	3.08		
<i>Elaeagnus pungens</i>	Site B		2090				5.64	44.5	tr	7.45		
<i>Eriobotrya japonica</i>	Site A	tr	265		tr	3.92						
<i>Eurya japonica</i>	Site C	tr	299	46	tr	3.81	5.07	8.6	tr	tr	1370	1180
<i>Eurya japonica</i>	Site A	tr	575		tr	5.37						
<i>Eurya japonica</i>	Site A	tr	811		tr	8.43						
<i>Eurya japonica</i>	Site A	tr	502	54.4	tr	5.59	1.56	9.36	tr		7180	11700
<i>Eurya japonica</i>	Site B		786				3.56	8.35	tr	tr		
<i>Eurya japonica</i>	Site B		730				3.82	10.3	tr	tr		
<i>Evodiopanax innovans</i>	Site B	tr	1150	80.5	tr	2.07	5.73	277	6.55	0.69		
<i>Evodiopanax innovans</i>	Site B	tr	214	77	tr	1.37	6.19	173	5.66	0.44		
<i>Evodiopanax innovans</i>	Site B	tr	577	75.5	tr	2.48	3.84	331	14	0.46		
<i>Evodiopanax innovans</i>	Site B	tr	154	73.9	tr	2.6	5.27	219	4.89	1.03		
<i>Fagara mantchurica</i>	Site A		78	21.7	tr	1.69	2.2	7.8	tr		18800	3300
<i>Ficus erecta</i>	Site A	tr	197	151	tr	6.96	5.67	29.8	tr		20200	44200
<i>Gardenia jasminoidesw</i>	Site A	tr	37	48.9	tr	10.7	3.86	10.7	tr		12800	10300
<i>Gleichenia dichotoma</i>	Site A	tr	1420		tr	6.1						
<i>Gleichenia dichotoma</i>	Site A	tr	1790		tr	2.07						
<i>Gleichenia dichotoma</i>	Site A	tr	600	87.3	tr	3.17	4.83	26.8	tr		1170	
<i>Gleichenia glauca</i>	Site A	tr	1280		tr	21.1						
<i>Hamamelis obtusata</i>	Site C	1.69	128		tr	0.98	7.59	14.8	tr	2.85		
<i>Hamamelis japonica</i>	Site B	tr	443	72.4	tr	4.92	5.57	13.1	0.24	1.88		
<i>Hydrangea hirta</i>	Site C	tr	53	98.1	tr	5.24	9.2	57.5	tr	tr	2660	
<i>Hydrangea hirta</i>	Site B	tr	121	380	tr	11	6.31	30.1	tr	3.94		
<i>Ilex crenata</i>	Site C		295	60.4	tr	3.58	4.37	258	0.86	tr	3600	5800
<i>Ilex crenata</i>	Site B	0.78	1037				2.43	931	14.77	tr		
<i>Ilex crenata</i>	Site B	tr	3080	98.8	tr	5.11	5.33	1580	29.3	0.47		
<i>Ilex crenata</i>	Site B	tr	2780	83	tr	11.4	5.04	459	16.2	1.49		
<i>Ilex crenata</i>	Site B		955				4.65	656	7.42	tr		
<i>Ilex crenata</i>	Site B	tr	1270	74.4	tr	6.13	3.6	908	23.5	1.32		
<i>Ilex crenata</i>	Site B		3340				5.79	828	10.4	tr		
<i>Illicium relucium</i>	Site C	tr	93	34.9	tr	4.43	1.73	9.19	tr	tr	2250	1060
<i>Illicium relucium</i>	Site B		192				3.5	13.7	tr	tr		
<i>Illicium relucium</i>	Site B		312				1.96	9.45	tr	tr		
<i>Illicium relucium</i>	Site B		236				3.14	14.2	tr	tr		
<i>Lespedeza bicolor</i>	Site A	tr	45	69	tr	11.9	6.28	11.7	tr		5750	3030
<i>Ligularia tussilaginea</i>	Site A	tr	50		tr	24.6						
<i>Ligularia tussilaginea</i>	Site A	tr	76		tr	18.5						
<i>Ligustrum japonicum</i>	Site A	tr	42		tr	11.7						
<i>Ligustrum japonicum</i>	Site A	tr	38	31.3	tr	5.53	2.95	25.8	tr		5310	3000
<i>Ligustrum japonicum</i>	Site A	tr	102			1.05						
<i>Ligustrum japonicum</i>	Site B		145				5.17	74.2	tr	tr		
<i>Litsea glauca</i>	Site C	tr	278	76.5	tr	1.51	4.91	29.5	tr	tr	856	3530

plant species	sampling site	Cr	Mn	Fe	Co	Ni	Cu	Zn	Cd	Pb	Mg	Ca
<i>Litsea glauca</i>	Site A	tr	166		tr	20.3						
<i>Litsea glauca</i>	Site B		145				4.87	44.2	0.92	tr		
<i>Litsea glauca</i>	Site B		220				5.69	47.6	0.88	2.95		
<i>Litsea glauca</i>	Site B		185				7.62	30.8	1.79	tr		
<i>Lycopodium serratum</i>	Site A	2.28	47		tr	26.5						
<i>Machilus Thunbergii</i>	Site A	tr	159	70.3	tr	15.5	9.84	36.8	tr		2880	1730
<i>Maesa japonica</i>	Site A		19		tr	12.66						
<i>Mallotus japonicus</i>	Site C	2.42	234		tr	0.91	7.26	40.1	tr	2.79		
<i>Mallotus japonicus</i>	Site A		137	77.8	tr	8.76	5.81	23	tr		11200	9600
<i>Meliosma myriantha</i>	Site C	2.18	116		tr	0.39	6.03	13.3	tr	4.11		
<i>Miscanthus sinensis</i>	Site A	tr	225		tr	2.08						
<i>Myrica rubra</i>	Site A	tr	706		tr	5.07						
<i>Myrica rubra</i>	Site A	tr	601		tr	14.5						
<i>Oenanthe Stolonifera</i>	Site A	tr	419		tr	18.9						
<i>Ophiopogon japonicus</i>	Site A	tr	46		tr	14.7						
<i>Orixa japonica</i>	Site C	tr	38	48.5	tr	3.06	10.6	26.9	tr	tr	3450	9390
<i>Orixa japonica</i>	Site B	tr	64	92	tr	1.99	6.5	26	0.15	1.49		
<i>Osmanthus ilicifolius</i>	Site C	tr	42	31.5	tr	0.59	2.58	26.3	tr	tr	479	10200
<i>Osmanthus ilicifolius</i>	Site A	tr	197		tr	34.4						
<i>Osmanthus ilicifolius</i>	Site A	tr	79		tr	23.8						
<i>Osmanthus ilicifolius</i>	Site A	tr	257	169	tr	26.1	4.1	32.1	tr		8510	7280
<i>Ostrya japonica</i>	Site C	2.35	328		tr	2.71	7.61	12.4	tr	3.31		
<i>Parabenzoin trilobum</i>	Site B	tr	835	99.2	tr	2.89	11.6	105	6.22	1.27		
<i>Pieris japonica</i>	Site C		219	71.8	tr	2.17	4.48	13.7	tr	tr	730	7700
<i>Pieris japonica</i>	Site A	3.55	2491		tr	22.41						
<i>Pieris japonica</i>	Site A		1208		tr	15.18						
<i>Pieris japonica</i>	Site A		153	84.4	tr	12.15	3.2	13.8	0.99		6010	8500
<i>Pieris japonica</i>	Site B		962				6.84	33.4	1.71	tr		
<i>Pieris japonica</i>	Site B		608				4.83	33.8	0.78	tr		
<i>Pieris japonica</i>	Site B		1475				4.05	15.5	tr	tr		
<i>Phalaris arundinacea</i>	Site A	tr	87		tr	3.17						
<i>Rhododendron macrosepalum</i>	Site A	4.16	947		tr	6.51						
<i>Phyllostachys bambusoides</i>	Site A	tr	13		tr	5.13						
<i>Pinus Thunbergii</i>	Site A	tr	66		tr	22.9						
<i>Pleioblastus Simonii</i>	Site A	tr	71		tr	2.82						
<i>Podocarpus macrophylla</i>	Site A	tr	26		tr	6.46						
<i>Podocarpus macrophylla</i>	Site A	tr			tr	2.32						
<i>Populus Sieboldi</i>	Site C	tr	73	38.8	tr	0.52	5.9	69.3	tr	tr	2680	4710
<i>Quercus acutissima</i>	Site A	tr	768	55.3	tr	6.07	3.95	11	tr		4420	5270
<i>Quercus crispula</i>	Site C	0.82	159		tr	1.3	2.9	16.6	tr	1.62		
<i>Quercus phillyraeoides</i>	Site A	tr	1110		tr	10.1						
<i>Quercus phillyraeoides</i>	Site A	tr	751		tr	17.2						
<i>Quercus phillyraeoides</i>	Site A	tr	777		tr	14.7						
<i>Quercus phillyraeoides</i>	Site A	tr	381	140	tr	10.8	3.06	10.5	tr		6130	8350
<i>Quercus serrata</i>	Site A	tr	386	79.4	tr	7.25	3.31	13.1	tr		6930	6530
<i>Quercus serrata</i>	Site B	tr	309	59.4	tr	6.61	8.97	21.7	tr	1.38		
<i>Rapanaea neriifolia</i>	Site A	tr	46		tr	6.25						
<i>Rapanaea neriifolia</i>	Site A	tr	66		tr	14.5						
<i>Rhododendron Kaempferi</i>	Site B	tr	181	127	tr	5.7	5.99	35.4	3.26	4.69		

plant species	sampling site	Cr	Mn	Fe	Co	Ni	Cu	Zn	Cd	Pb	Mg	Ca
<i>Rhododendron quinquefolium</i>	Site C	tr	33	92	tr	3.22	7.12	20.8	tr	tr	2490	1650
<i>Rhododendron reticulatum</i>	Site C	tr	68	79.3	tr	5.25	6.26	21.2	tr	tr	4460	5150
<i>Rhododendron sanctum</i>	Site A	tr	315	162	1.45	19.6	2.08	11.2	0.42		10300	6870
<i>Rosa multiflora</i>	Site A	tr	27		tr	16.2						
<i>Rubus Burgeri</i>	Site A	tr	677		tr	29.4						
<i>Rubus Burgeri</i>	Site A	tr	161		tr	22.5						
<i>Rubus crataegifolius</i>	Site A	tr	80		tr	18.8						
<i>Salix integra</i>	Site C	2.37	26		tr	1.52	5.51	75.2	0.28	3.34		
<i>Sapium japonicum</i>	Site C	tr	38	41.9	tr	0.83	7.02	23	tr	tr	2170	2070
<i>Skimmia japonica</i>	Site B		44				3.81	37.7	2.78	tr		
<i>Skimmia japonica</i>	Site B		86				3.62	59.1	1	tr		
<i>Solidago altissima</i>	Site A	tr	55		tr	23.3						
<i>Sonchus oleraceus</i>	Site A	tr	53		tr	3.96						
<i>Sphenomeris chusana</i>	Site A	tr	54		tr	5.09						
<i>Sphenomeris chusana</i>	Site A	tr	100		tr	8.12						
<i>Stephanotis japonica</i>	Site A	tr	214		tr	35.1						
<i>Stephanotis japonica</i>	Site A	tr	56		tr	13.6						
<i>Stewartia monadelpha</i>	Site C	2.93	315		tr	3.88	3.47	15	tr	3.39		
<i>Torreya nucifera</i>	Site C	tr	33	45		1.82	5.17	14.9	tr	tr	1050	5310
<i>Torreya nucifera</i>	Site B		681				4.93	19.7	tr	tr		
<i>Trachycarpus excelsa</i>	Site A	tr	32		tr	13.5						
<i>Verbena phlogiflora</i>	Site C	2.63	77		tr	1.59	9.08	27	tr	3.91		
<i>Verbena phlogiflora</i>	Site A	tr	23	133	tr	10.7	11.1	18.4	tr		6520	5640
<i>Viburnum tomentosum</i>	Site C	tr	57	59.4	tr	0.25	7.32	15	tr	tr	3930	6080
<i>Wistaria floribunda</i>	Site C	1.52	408		tr	1.56	8.15	34	tr	3.13		
<i>Wistaria floribunda</i>	Site A	tr	281	88.9	tr	7.08	3.6	15.1	tr		7410	9380

tr; trace no datum; not determined

Site A, Toba city Site B, Mt.Fujiwara Site C, University forest

Table 3 Differences of heavy metal concentrations in the leaves of *Clethra barbinervis* sampled from three sites
mg kg⁻¹

Sampling Site	Cr	Mn	Fe	Co	Ni	Cu	Zn	Cd	Pb	Mg	Ca
SiteA	2.06	955	151	108	212	5.42	96	0.66	2.91	14100	5020
SiteB	0.22	1270	92	41	24.9	7.24	241	8.71	2.27		
SiteC	1.84	564	50	12	7.6	6.63	192	0.9	2.97	4150	1210

Site A, Toba city Site B, Mt. Fujiwara Site C, University forest

At Site C, Mn, Co, and Zn concentrations in *C. barbinervis* were higher than those in soil (Mn: 564 mg kg⁻¹ in *C. barbinervis* and 341 mg kg⁻¹ in soil; Co: 12.1 mg kg⁻¹ in *C. barbinervis* and 5.22 mg kg⁻¹ in soil; Zn: 192 mg kg⁻¹ in *C. barbinervis* and 102, Table 1 and 3). *C. barbinervis* also accumulated Cd (0.9/0.0). In contrast to *C. sciadophylloides*, *C. barbinervis* seems to absorb many kinds of heavy metals from soil but its ability to transfer Mn from soil to plant was lower than that of *C. sciadophylloides*.

As shown in Table 3, average concentrations of Co, Ni, and Mg in the leaves of *C. barbinervis* were higher in leaves collected at Site A; and Zn and Cd concentrations were higher in leaves collected in Site B than at other areas. These trends may have resulted from the concentration of each heavy metal in soil at these sites. Ni concentration in *C. barbinervis* collected at Site A was highest but did not meet or exceed the criterion of hyperaccumulation by Brooks¹⁾ (1000 mg kg⁻¹). The concentrations of Zn and Cd are highest

in *Ilex crenata* from Site B (Table 2) but the concentrations did not meet or exceed the criteria by Brooks¹⁾, 10,000 and 100 mg kg⁻¹ respectively.

The concentration of heavy metals in wild plants varies according to plant species and further study is required to search for more hyperaccumulator plants in Japan.

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三重県下三地点における植物葉中の重金属元素濃度

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要 旨

重金属回収に用いる植物を探索するため、三重県下三地点における木本、草本植物の葉の重金属元素濃度を測定した。その結果、重金属濃度は植物の種によって大きく変動することが明らかになった。同じ地点の同じ植物種であっても濃度は変動し、また、地点が異なる同じ植物種の場合には、その地点における土壌の当該重金属濃度に応じて葉中の濃度が大きく変動した。コシアブラ (*Changeopanax sciadophylloides*) は Brooks の定義による超集積性植物であること、マンガンのみを特異的に集積し他の重金属は集積しないことが確認された。また、リョウブ (*Clethra barbinervis*) は多種類の重金属元素を高濃度に集積するが、土壌からの移行係数はコシアブラによるマンガン吸収の場合に比べて小さかった。